

# OPTIMIZATION OF THE DERIVATIZATION OF SIX PROBE METABOLITES FOR THE IN VITRO DETERMINATION OF CYTOCHROME P450 ACTIVITY IN CHILDREN WITH HEPATIC IMPAIRMENT.

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## 1. Introduction

In previously published studies, reduced cytochrome P450 enzyme activity was detected in adult patients with hepatic impairment. The activity of this essential enzyme system in children with severe hepatic dysfunction, has not yet been assessed. For an in vitro determination of the activity of the six most important isoforms, 1A2, 2C9, 2C19, 2D6, 2E1 and 3A4, hepatic microsomes can be incubated with specific probe substrates that are selectively metabolized by one of the isoforms.

## 2. Aim

Enzyme activity is expressed as amount of metabolite formed per milligram of protein and per minute. To quantify the amount of metabolite formed, a quantification method was developed using HPLC-MS/MS (Micromass, Quattro II). This method had two major limitations: insufficient sensitivity was seen for the detection of some of the metabolites, and some metabolites could only be detected in the negative electrospray ionization mode. To overcome these limitations, derivatization of the molecules was considered.

## 3. Method optimization: HPLC-MS/MS

### Probe substrates and metabolites

CYP	Substrate	Concentration (μM)	Metabolite
1A2	Phenacetin (PH)	50	Acetaminophen (AP)
2C9	Tolbutamide (TB)	100	4-OH-tolbutamide (HTB)
2C19	S-mephenytoin (ME)	100	4'-OH-mephenytoin (HME)
2D6	Dextromethorphan (DM)	5	Dextrorphan (DX)
2E1	Chlorzoxazone (CZ)	50	6-OH-chlorzoxazone (HCZ)
3A4	Midazolam (MDZ)	5	1-OH-midazolam (HMDZ)

Table 1: Probe substrates and metabolites

### Sample matrix :

- 0.25 mg/ml microsomal protein
- 0.2 M phosphate buffer (pH 7.4)
- 1 mM NADPH
- 1.15% KCl
- spiked metabolites

### MS/MS Method (Micromass Quattro II)

- Capillary voltage: 3.6 kV
- Gas flow: drying gas 450 L/h and nebulising gas 6 L/h
- Source temperature : 120°C

### HPLC Method (Kontron)

- Luna C18 Column 3 μm 50 x 2.0 mm, 100Å (Phenomenex) and Alltima C18 Pre-column 5 μm 7.5 x 2.1 mm (Grace)
- Eluent A: Water + 0.1% formic acid
- Eluent B: Acetonitrile + 0.1% formic acid
- Flow: 200 μl/min
- Partial loop injection (10 μl)

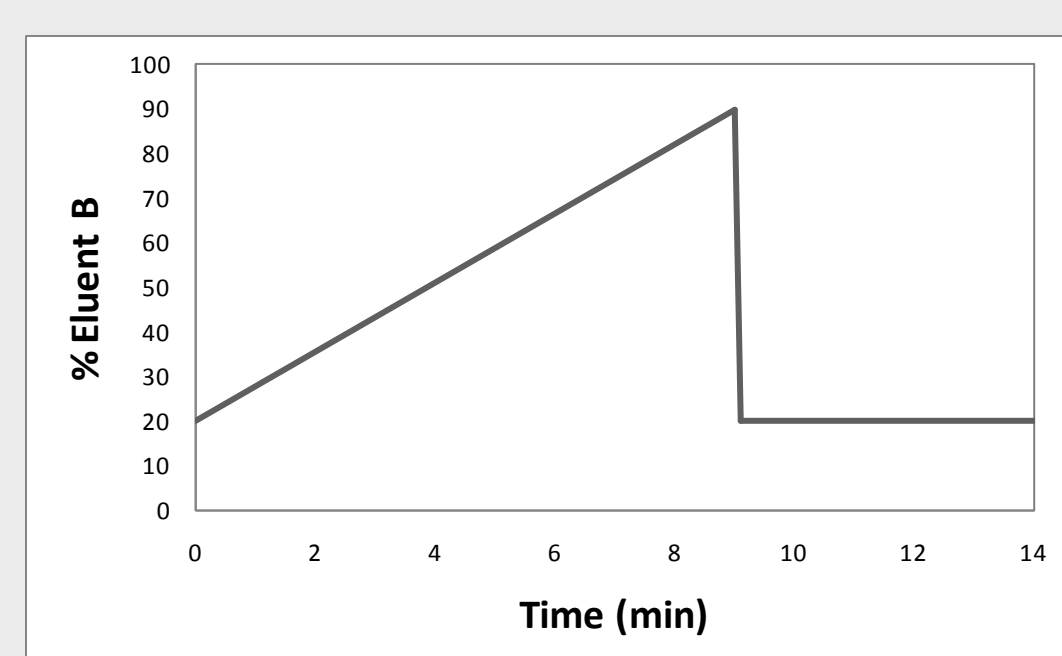


Figure 1: HPLC gradient profile

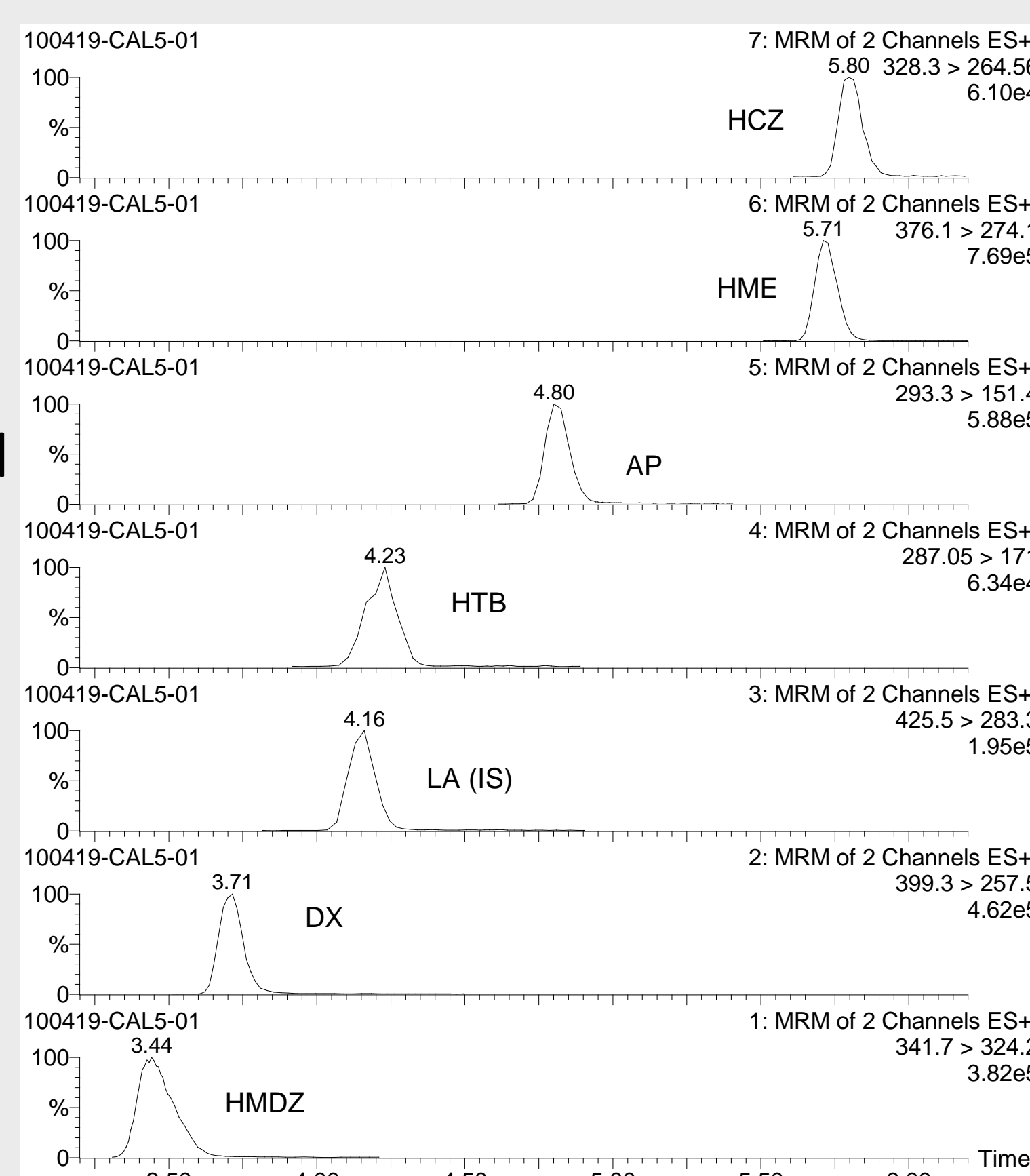


Figure 2: Chromatogram of a calibrator obtained with the final method

## 4. Method optimization: derivatization

- **Derivatization reagent:** Pyridine-3-sulfonyl chloride hydrochloride (PS) versus 4-(1H-pyrazol-1-yl)benzenesulfonyl chloride (PBS): derivatization of the aromatic OH-function (in semi-aqueous reaction environment)

PS was chosen for the following reasons:

- good fragmentation pattern
- shorter retention times

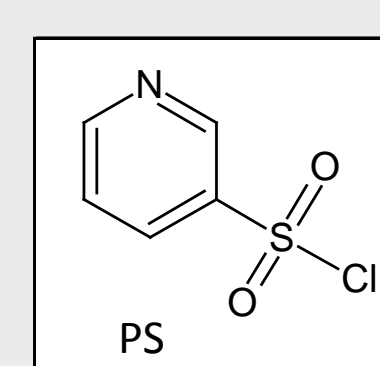


Figure 4: Structure of derivatization reagent

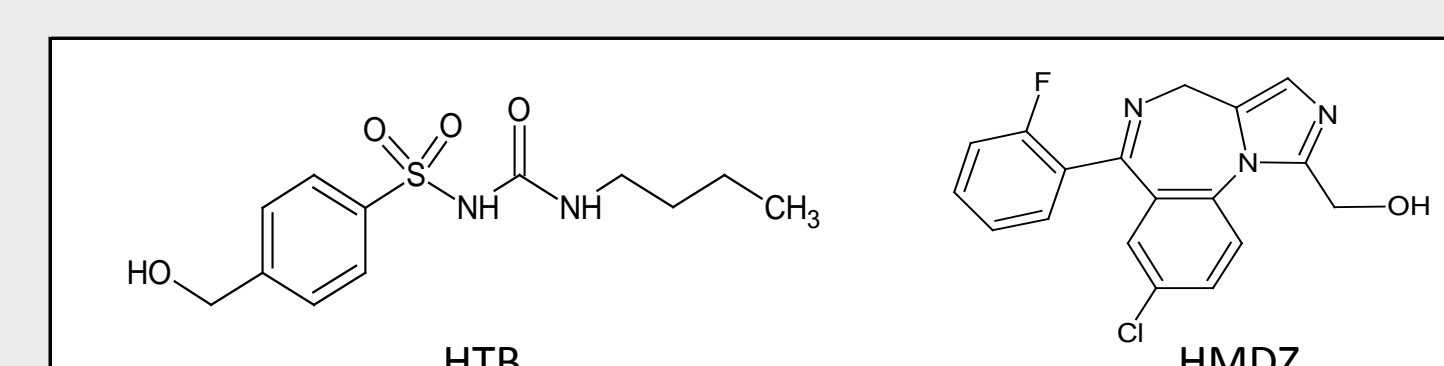


Figure 3: Structure of the two underivatized metabolites

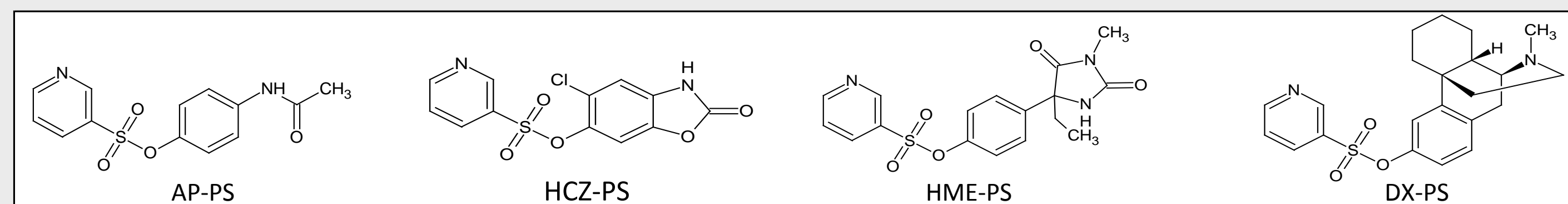


Figure 5: Structures of the four derivatized metabolites

- **Other parameters optimized and interpreted based on sensitivity:**

Temp (°C), heat block	40 - 55 - 60 - 70 - 80 - 90
Time (min)	5 - 10 - 15 - 20 - 30
pH	9 - 9,5
Cool down time (min)	room temp 10 - ice 5 - ice 10
Amount PS (μl)	50 - 60 - 70
Microwave 850 Watt (sec)	10 - 20 - 30 - 40

### Final derivatization protocol:

- 200 μl sample
- + 10 μl 1.75 M NaOH
- + 10 μl internal standard (Levallorphan 1.25 μg/ml)
- + 70 μl 1mg/ml PS in acetonitrile
- vortex mixing
- derivatization reaction for 20 seconds in microwave (850 Watt)
- vortex and cool on ice for 10 min
- transfer to 250 μl insert for injection (note: no concentration step !)

	Ionization mode	Precursor (m/z)	Quantifier (m/z)	Qualifier (m/z)	Cone voltage (V)	Collision energy (eV)	Retention time (min)	LOQ (ng/ml)
AP-PS	ESI +	293,30	151,40	109,10	40	18	4,80	5,6
HTB	ESI +	287,05	171,00	188,00	35	18	4,23	25,0
HME-PS	ESI +	376,10	274,10	132,10	45	23	5,71	6,9
DX-PS	ESI +	399,30	257,50	199,02	50	30	3,71	1,1
HCZ-PS	ESI +	328,30	364,56	144,25	35	17	5,80	128,1
HMDZ	ESI +	341,70	324,20	203,20	40	20	3,44	2,3

Table 2: Individual parameters for the six (derivatized) probe metabolites

## 5. Conclusion

This quantification method shows clear improvement compared to the previously developed method: only positive electrospray ionization is necessary, and the limits of quantification of all metabolites are in the range needed for the particular preclinical pharmacokinetics application. The procedure is quick and easy to perform without various extraction/re-concentration steps. This method is now in the process of full validation.