

Correlation between endoreduplication and metabolism in *Arabidopsis thaliana*: an LC-MS based metabolomics approach

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- Introduction
- Analytical approach
- Biological approach
- Analytical performance
- Trial LC-MS metabolome comparison
- Future?





Introduction



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Introduction

- Endoreduplication?
 - No mitosis and/or cytokinesis
 between successive rounds of
 DNA replication
 - DNA content of cell is doubled with every round of DNA replication
 - o Formation of cells with DNA ploidy level of 2C, 4C, 8C, 16C, 32C...

=> polyteny





Introduction

- Function of endoreduplication?
 - o *Arabidopsis thaliana*: present in practically all tissues
 - o <u>Hypotheses</u>:
 - Endoreduplication ~ fast life cycle and improved production stability
 - o Buffering of mutations to preserve functional copies of the genome (Inzé et al., 2006)
 - o Micro-array experiments: upregulation of genes involved in nitrogen assimilation and metabolism (Vlieghe et al., 2003)
 - o Level of endoreduplication ~ developmental stage



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Analytical approach



Endoreduplication and LC-MS based metabolomics



Analytical approach

- LC-MS analysis (J. Sep. Sci 2007, 30, 2002-2011)
 - Alliance 2690 LC system (Waters, Milford, MA)
 - Eluent A: $H_2O + 0.1\%$ FA
 - Eluent B: AcN/H₂O 90/10 + 0.1% FA
 - Atlantis dC18 (2.1 x 150mm) column
 - QTOF micro (Waters, Milford, MA)
 - Lockspray





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Biological approach







Biological approach

Flow cytometry:
 DNA content
 ³⁰
 ⁴⁰
 ⁷⁰
 ploidy level





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Analytical performance

- LC-MS precision
 - Variability of the LC/MS-tool through consecutive injections of the same extract
 - 25 m/z-tR combinations were randomly chosen out of the chromatogram, covering the tR-range and the m/z-range of our LC/MS tool and peak area deviation was evaluated
 - Primary leaf pairs extract (n=5)



8.88 ± 5.16%



- Analytical variation
 - includes the extra variability of the pre-LC/MS procedure
 - Pooled primary leaf pairs extract, 5 samples











- Biological variation
 - variability in plant growth conditions, development etc. on sample plates within primary leaf pairs of wild-type *Arabidopsis thaliana*







Analytical performance



Endoreduplication and LC-MS based metabolomics



Analytical performance

- Absolute recovery (AR) & Process efficiency (PE)
- Out of the broad range of *Arabidopsis thaliana* metabolites, we selected a group of 16 metabolites, representative for different chemical compound classes
- Analysis of 6 extracts with metabolites spiked before extraction (PRE), 6 extracts with metabolites spiked after extraction (POST), 6 blank extracts (BLANC) and 6 pure metabolite mixtures (in solvent, PURE).

AR = % [peak area PRE / peak area POST]

PE = % [(peak area PRE - peak area BLANC) / peak area PURE]





294.7% 100% 50% metabolite 0% cytidine adenine zeatine leucine phenylalanine S-adenosylmethionine UMP gibberellic acid jasmonic acid abscisic acid spermidine chorismic acid coumaric acid epibrassinolide ADP-glucose actos -50% □ absolute recovery (AR) process efficiency -100%

Analytical performance

Endoreduplication and LC-MS based metabolomics





- Matrix effect (ME)
 - o Post-column infusion of leu-enk
 - o Blank run vs plant extract





- Matrix effect (ME)
 - o Post-extraction addition
 - o ME = 100% %[(peak area POST- peak area BLANC) / peak area PURE]



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Primary leaf pairs of:





- Analysis in both + and ESI mode
- *m/z*-range 100-1000
- Data processing
 - o MarkerLynx
 - o Simca-P





- PLS-DA (supervised)
- Negative ESI mode:
 - 15 observations
 - 822 variables
 - $R^2 X = 0.836$
 - $R^2 Y = 0.986$





- PLS-DA (supervised)
- Positive ESI mode
 - 15 observations
 - 2034 variables
 - $R^2 X = 0.730$
 - $R^2Y = 0.977$













Endoreduplication and LC-MS based metabolomics



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Future

- LC-MS metabolome comparison of a large batch of different genotypes
- QC inclusion (Gika et al., 2007)
- HILIC approach for polar metabolite fraction
- Additional GC-MS analysis
- FT-MS analysis towards identification

Thank you !